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A technique of hydride generation (HG) with subsequent trapping, CT) under liquid nitrogen was developed in the early 1970's to improve limits of detection for elements like arsenic and selenium. However, the unique features of this technique make it attractive even for modern analytical chemistry.

DART-MS STUDY OF SELENIUM HYDRIDE AND METHYLATED ARSANES.

For studies of processes taking place e.g. in hydride generation or atomization, CT can effectively separate the species of interest from other compounds and also from accompanying ballast from HGspray droplets, vapors and hydrogen. Presented examples are mass spectra of generated selenium hydride, arsane, mono-, di- and trimethylarsane from a DART ionization source coupled with an LTQ-Orbitrap mass spectrometer.

HG-CT: **HG-CT- DART- LTQ-Orbitrap MS setup**

ICP-MS ANALYSIS OF TRACE ARSENIC SPECIES IN WATER REFERENCE MATERIALS

HG-CT offers the possibility of preconcentration of analyte from large sample volumes, as well as the ability to separate individual species. This is ideal for speciation analysis at ultratrace levels, which is one of the current challenges of elemental analytical chemistry. For many matrices, minimal sample pretreatment is required, which reduces sample dilution and risk of contamination.

Se: 2 ppm in 1 M HCl 0.5% NaBH₄ in 0.4% KOH As species: 1 ppm in 1 M HCl 1% NaBH₄ in 0.1% KOH Flow rates: 1mL/min Carrier gas He 100 mL/min

Sample volume: 0.5 mL

DART settings: Needle electrode voltage 3500V He DART flow rate 3.5 L/min Discharge & grid electrodes 0V

LTQ-Orbitrap MS settings: Resolution 15 000

Capillary temperature 100°C Capillary voltage -35V (negative mode); +65V (positive mode) Tube lens voltage -45V (negative mode); +100V (positive mode)



CONCLUSIONS:

Spectra in both positive and negative ion mode contain mainly the oxidized ions, with the hydride ion not present (As) or present as a minor ion (Se).

Losses of one or more methyl groups are observed for methyl substituted arsanes.

Numerous ions contain multiple As or Se atoms, which do not reflect original species but originate in the ion source. This is more pronounced in HG-CT spectra compared to HG without cryotrap.

Reactions in the ion source must be taken into account in the interpretation of DART spectra for the mechanistic studies of HG.

RFS	SULTS:	Carrier gas: Ar 0.6 L/min		
SeH ₂ , negative mode:	Dilution gas: 0.55 L/min 1 U-tube precool & sample inject 30 -			
Mass spectra taken with a cryotrap, with cryotrap and 15 mL/min	in Mass spectra taken with a cryotrap.	75 Consider cell. He 3.5 mi/min 2 Hydride generation 75 C 75As 3 HG system gas flush 85		
H_2 added to simulate H_2 produced from the generator, and directly from hydride generator without a cryotrap.	tly Maca Tantativa Dalta AcH MAcH DMAcH TMAc	IS 125Te 100 ng/mL, 4 U-tube heat & data acquisition 80 -		
	Formula mmu	nebulized <u>5 U-tube drying & waste liquid removal 85 -</u>		
Mass Tentative Delta HG-CT HG-CT HG only	106.9133 O2 As 2.64 10% 6% 5% 3% 122.9080 O3 As 2.82 9% 13% 13% 8%			
80.9263 H Se 2.52 10% 11% 8% 05.0400 0.0 0.57 4% 4% 4%	123.9159 H O3 As 2.79 3% 5% 4% 124.9247 H2 O3 As 2.72 19% 14% 7% 4%	For complete method description see: Matoušek et al, JAAS., 28 (2013),		
95.9132 0 Se 2.57 1% 2% 1% 111.9088 02 Se 2.66 30% 28% 32%	136.9612 C2 H6 O2 As 2.88 65% 100% 137.9634 C H3 O3 As 2.90 4% 4%			
128.9115 H O3 Se 2.78 67% 41% 44% 144.9061 H O4 Se 2.84 100% 100% 100%	138.9395 C H4 O3 As 2.87 31% 100% 61% 139.9109 C H4 O3 As 2.84 11% 9% 6%	RESULIS:		
157.9013 O4 N Se 2.79 8% 5% 5% 161.9098 H2 O5 Se 2.89 8% 6% 7%	140.9191 H2 O4 As 2.86 100% 100% 73% 32% 153.9142 H O4 N As 2.81 14% 13% 8%	Arsenic speciation in the new certified reference materials of river, sea and		
173.8969 O5 N Se 2.99 12% 7% 4%	154.9350 C H4 O4 As 2.87 4% 8% 5% 155.9056 H O5 As 2.83 7% 6% 4%	na/L (sd) na/L (sd) na/L (sd) na/L (sd)		
192.8340 H 02 Se2 3.25 38% 50% 13% 208.8276 H 03 Se2 3.29 4% 4% 1%	156.9135 H2 O5 As 2.99 5% 7% 4%	SLRS-6 Riverine water 362 (4.1) 75 (2.3) 73 (2.8) 21 (1.1)		
225.8314 H2 O4 Se2 3.36 3% 6% 1% 240.8181 H O5 Se2 3.41 8% 4%	158.9302 H4 O5 As 2.86 7% 6% 4%	NASS-7 Seawater 846 (4.6) 19 (0.4) 193 (1.9) 13 (0.3) CASS 6 Seawater 400 (7.2) 20 (0.8) 400 (2.6) 24 (0.2)		
256.8120 H O6 Se2 3.51 3% 3% 272.7510 H O2 Se3 3.82 3% 3%	167.9433 C2 H5 O4 As 3.03 7% 15% 167.0400 C2 H5 O4 As 3.03 7% 15%	CASS-6 Seawater 499 (7.2) 20 (0.8) 199 (2.6) 24 (0.3) AQUA-1 Drinking water 124 (2.0) 36 (0.9) 28 (1.9) 13 (0.3)		
Most abundant ion count: 5.6E+05 5.8E+05 1.1E+06	168.9509 C2 H6 O4 As 3.02 5% 14% 169.9584 C2 H7 O4 As 3.02 28% 46%			
	171.9384 C H5 O5 As,H3 O5 3.02 12% 12% 33% 18% 172.9376 C2 H7 O2 As Cl 3.05 9% 11%			
	173.9177 H3 O6 As 2.94 23% 23% 14% 6% 174.9167 C H5 O3 As Cl 2.90 22% 11%	The reference materials were analyzed after prereduction with 0.1g solid L-cysteine hyd DMAs) or without prereduction (TMAsO). Triplicate analyses of 3 subsamples of 3 ba		
	176.8962 H3 O4 As Cl 2.94 11% 4% 13% 8% 182.9658 C3 H8 O4 As 2.95 4%	point calibration for all species prepared in 0.025% HNO ₃ .		
Arsines, positive mode: Mass spectra taken with a cryotrop	183.9609 C2 H7 O4 N As 3.05 22% 33% 184.9452 C2 H6 O5 As 3.17			
ινία το τη	185.9412 C H5 O5 N As 3.05 9% 29% 13%	⁶⁰⁰⁰⁰⁰ J		
Mass Tentative Delta AcH MAcH DMAcH TMAc	187.9204 H3 O6 N As 3.01 13% 14% 10% 4% 106.0826 C4 140 O4 Ac 2.05 13% 14% 10% 4%	50000 - BLANK materials.		
Formula mmu	196.9826 C4 H10 O4 As 3.05 7% 198.9609 C3 H8 O5 As 3.02 3%			
88.9358 C H2 As -0.71 1% 90.9153 O As -0.71 3% 3%	199.9570 C2 H7 O5 N As 3.10 7% 12% 3% 201.9348 C2 H7 O6 As 3.07 8% 29% 12%	iAs iAs		
90.9516 C H4 As -0.72 1% 100.9361 C2 H2 As -0.63 1% 4%	202.9206 C H4 O7 As 3.14 4% 4% 4% 203.9157 H3 O7 N As 3.10 18% 16% 21% 12%	₩ 300000 - iAs		
102.9518 C2 H4 As -0.58 8% 104.9675 C2 H6 As -0.55 11% 10%	8% 214.8321 H O4 As2 3.00 4% 6%	200000 -		
105.9627 C H5 N As -0.58 4% 106 9468 C H4 O As -0.44 17% 10% 49	229.8194 O5 As2 3.18 3% 4% 230.8283 H O5 As2 3.21 15% 24% 8%	100000 - <u>MAs</u>		
108.9624 C H6 O As -0.54 3%	232.8446 H3 O5 As2 3.27 9% 11% 2% 242.0012 C2 H0 O2 Ac2 3.20 9% 0%			
118.9832 C3 H8 As -0.44 49 100.0000 C3 H8 As -0.44 49	2% 242.9012 C3 H9 C3 AS2 3.30 9% 4% 244.8811 C2 H7 O4 As2 3.43 15% 51% 9% 4% 244.8811 C2 H7 O4 As2 3.43 15% 51% 9%	0 20 40 60 80 6 000 000		
120.9989 C3 H10 As -0.41 2% 7% 136.9943 C3 H10 O As -0.28 79%	7% 246.8589 C H5 05 As2 3.34 48% 43% 4% 9% 247.8318 H2 06 As2 3.35 4%	3 000 000		
137.9653 C2 H7 O2 As -0.32 2% 138.9728 C2 H8 O2 As -0.15 100% 100%	2% 248.8401 H3 O6 As2 3.39 32% 40% 15% 0% 258.8955 C3 H9 O4 As2 3.45 48% 32%	L-cysteine prereduced Not prereduced		
140.9526 C H6 O3 As -0.15 100% 4% 142.9317 H4 O4 As -0.28 100% 9%	4% 259.8676 C2 H6 O5 As2 2.99 7% 16% 260.8739 C2 H7 O5 As2 3.53 11% 72% 23%	pg/ml pg/ml pg/ml		
154.9683 C2 H8 O3 As -0.07 3% 3% 164.8660 C H3 As2 -0.11 13%	261.8476 C H4 O6 As2 3.37 19% 14% 262.8557 C H5 O6 As2 3.45 26% 47% 9%	Subsample 1 851 18.7 197 12.4		
168.9842 C3 H10 O3 As 0.17 12% 1% 178.8820 C2 H5 As2 0.19 1%	263.8276 H2 O7 As2 3.41 7% 9% 264.8342 H3 O7 As2 3.43 24% 30% 22% 4%	<u>852 18.6 196 12.7</u> 847 18.7 194 13.1 MAs		
194.9130 C3 H9 As2 -0.03 4% 6%	273.8707 C2 H6 O5 N As2 3.49 5%	Bottle 1 Subsample 2 847 18.6 194 13.3 600 000 852 18.7 195 13.1 600 000		
198.8352 H O3 As2 0.03 7% 2%	275.8989 C3 H10 O5 As2 3.39 3%	848 18.7 194 13.2 gg Subsemple 2 852 10.0 10.1 10.1		
210.9082 C3 H9 O AS2 0.21 4% 4% 4% 4% 4%	278.8868 C2 H9 O6 As2 3.43 8% 7%	Subsample 3 852 18.9 195 13.1 2400 000		
214.8666 C H5 O3 As2 0.11 2% 216.8457 H3 O4 As2 -0.05 6%	279.8570 H4 08 As2 3.47 3% 22% 280.8644 C H7 O7 As2 3.41 4% 5%	849 18.1 195 12.9 Subsample 1 852 18.1 194 12.9 200 000		
224.9603 C5 H15 As2 0.26 4% 226.9396 C4 H13 O As2 0.33 8% 6%	4% 281.8378 H4 O8 As2 3.47 5% 7% 6% 289.9151 C2 H6 O6 N As2 3.53 3% 4%	851 18.1 194 13.0 844 19.5 193 13.4		
228.8822 C2 H7 O3 As2 0.01 6% 230.8615 C H5 O4 As2 0.08 11%	291.8454 C H4 O7 N As2 3.55 4% 6% 309.8576 C H6 O8 N As2 3.64 3%	Bottle 2 Subsample 2 839 19.5 193 13.3		
232.8406 H3 O5 As2 -0.11 11% 232.8772 C H7 O4 As2 0.15 2%	311.8327 H4 O9 N As2 3.41 3% 3% 320.7743 C H4 O5 As3 3.69 8%	<u>839 19.5 192 13.2</u> 842 19.0 191 13.0		
234.8549 H5 O5 As2 -0.05 2% 242.9345 C4 H13 O2 As2 0.34	322.7541 H2 O6 As3 3.76 5% 9% 2% 325.8517 C H6 O9 N As2 3.69 4%	Subsample 3 849 19.0 193 13.2 848 18.9 195 13.1 4 000 000		
244.9140 C3 H11 O3 As2 0.50 12%	334.7911 C2 H6 O5 As3 4.03 9% 6% 336 7669 C H4 O6 As3 3.77 20% 0%	848 18.3 188 12.9 DMAs Subsample 1 845 18.9 10.2 12.9		
248.8722 C H7 O5 As2 0.25 6%	338.7460 H2 O7 As3 3.88 10% 19% 4% 250.7822 C2 H6 O6 Ac2 2.00 10% 10% 10%	Subsample i 645 16.6 193 12.6 842 18.7 192 13.1 3 000 000		
254.8114 C2 H6 As3 0.23 6%	350.7623 02 F0 06 AS3 3.90 8% 12% 352.7614 C H4 O7 As3 3.91 19% 16%	847 18.4 192 13.4 Bottle 3 Subsample 2 838 18.2 192 13.4 <u> <u> </u></u>		
256.9503 C5 H15 O2 As2 0.41 209 258.9297 C4 H13 O3 As2 0.61 12% 489	0% 354.7420 H2 O8 As3 4.17 9% 13% 5% 8% 356.7598 H4 O8 As3 4.20 5%	838 18.1 192 13.3 843 18.9 192 13.1		
260.9091 C3 H11 O4 As2 0.68 5% 262.8879 C2 H9 O5 As2 0.27 4%	5% 366.8141 C3 H10 O6 As3 3.79 16% 368.7967 C2 H8 O7 As3 4.08 10% 19%	Subsample 3 842 19.1 192 13.1 1000 000 1000 000 1000 000 1000 000 1000 000		
264.8672 C H7 O6 As2 0.31 2% 266.8464 H5 O7 As2 0.24 3%	370.7738 C H6 O8 As3 4.00 9% 7% 372.7560 H4 O9 As3 4.32 3% 4%	846 19.1 193 13.2 Average 846 18.7 193 13.1		
270.8428 C3 H10 As3 0.28 5% 286.8378 C3 H10 O As3 0.42 2%	382.8134 C3 H10 O7 As3 4.18 6% 384.7904 C2 H8 O8 As3 4.03 5% 12%	RSD 0.5% 2.2% 1.0% 2.2%		
332.8440 C4 H12 O3 As3 1.08 1%	386.7712 C H6 O9 As3 4.39 4% 7% 400.8209 C3 H12 O8 As3 4.26 5%			
338.7812 C H6 O6 As3 0.54 2% 244.7567 C2 H0 Ac4 0.20 2%	402.7987 C2 H10 O9 As3 4.12 5% 404.7780 C H8 O10 As2 4.42 0%			
344.7507 03 H9 AS4 0.39 3% 348.8388 C4 H12 O4 As3 0.98 2%	404.7709 0 10 AS3 4.13 3% 444.6902 C H5 O8 As4 4.78 4% 450 7040 00 H7 00 A 4 0.45			
364.8701 C5 H16 O4 As3 1.02 2% Most abundant ion count: 1.5E+06 7.0E+06 1.7E+07 1.5E+07	-07 <u>458.7016</u> C2 H7 O8 As4 3.45 <u>3%</u> -07 Most abundant ion count: 9.9E+05 9.6E+05 3.8E+05 5.6E+05	ACKNOWLEDGEMENT. This study was supported by Czech Scie		

and methylated arsenic species is snown as an ction achieved with HG-CT coupled to quadrupo



ICP-MS

Flow rate: 1mL/min

HG-CT

Agilent 7700x with ISIS unit Power: 1600 W Carrier gas: Ar 0.6 L/min	Step	
Dilution gas: 0.55 L/min	1	U-tube precool & sample inject
Collision cell: He 3.5 ml/min	2	Hydride generation
75As	3	HG system gas flush
IS 125Te 100 ng/mL,	4	U-tube heat & data acquisition
nebulized	5	U-tube drving & waste liquid remov

	Time Pum		nps	U-tube	U-tube		
	S	1	2	Cooled	Heating		
ool & sample inject	30	-	ON	ON	-	-	
neration	75	ON	-	ON	-		
gas flush	85	-	-	ON	-		
& data acquisition	80	-	-	-	ON		
ng & waste liquid removal	85	-	ON	-	ON		

1456 - 1465.

nd drinking water

		iAs		MAs		DMAs		TMAsO		Sum of species	Certified As
		ng/L	(sd)	ng/L	(sd)	ng/L	(sd)	ng/L	(sd)	ng/L	ng/L
SLRS-6	Riverine water	362	(4.1)	75	(2.3)	73	(2.8)	21	(1.1)	531	570 ± 80
NASS-7	Seawater	846	(4.6)	19	(0.4)	193	(1.9)	13	(0.3)	1 071	1230 ± 60
CASS-6	Seawater	499	(7.2)	20	(0.8)	199	(2.6)	24	(0.3)	741	1040 ± 100
AQUA-1	Drinking water	124	(2.0)	36	(0.9)	28	(1.9)	13	(0.3)	201	not available yet

drochloride per 5 ml (iAs, MAs, ottles were analyzed against 5



as checked by matching ddition for all species in all



NASS-7 y = 6 517x + 129 066

0.025% HNO₃

0.025% HNO₃

300

y = 6342.8x + 20735

200

As, pg/mL

60

80

400

y = 6528.7x + 4900.2

As, pg/mL

NASS-7

y = 6 382x + 1 266 880

DDORT KVU: 68081/15

Foundation